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Temperature acclimation rate of aerobic scope and feeding metabolism in fishes: implications in a thermally extreme future

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Temperature acclimation may offset the increased energy expenditure (standard metabolic rate, SMR) and reduced scope for activity (aerobic scope, AS) predicted to occur with local and global warming in fishes and other ectotherms. Yet, the time course and mechanisms of this process is little understood. Acclimation dynamics of SMR, maximum metabolic rate, AS and the specific dynamic action of feeding (SDA) were determined in shorthorn sculpin (Myoxocephalus scorpius) after transfer from 10°C to 16°C. SMR increased in the first week by 82% reducing AS to 55% of initial values, while peak postprandial metabolism was initially greater. This meant that the estimated AS during peak SDA approached zero, constraining digestion and leaving little room for additional aerobic processes. After eight weeks at 16°C, SMR was restored, while AS and the estimated AS during peak SDA recovered partly. Collectively, this demonstrated a considerable capacity for metabolic thermal compensation, which should be better incorporated into future models on organismal responses to climate change. A mathematical model based on the empirical data suggested that phenotypes with fast acclimation rates may be favoured by natural selection as the accumulated energetic cost of a slow acclimation rate increases in a warmer future with exacerbated thermal variations.

1. Introduction

Temperature has profound effects on the performance and biogeography of ectothermic animals including fishes, partly through its effects on metabolism [1,2]. Increasing temperature results in an exponential increase in cellular respiration rate with a Q_{10} of approximately 2, which is met by a corresponding elevation of the whole-animal oxygen consumption rate (i.e. the standard metabolic rate; SMR) to meet aerobic energy requirements [3]. However, in the long term many species have a considerable capacity for physiological acclimation (i.e. reversible physiological plasticity) allowing them to compensate for the direct thermodynamic effects on metabolism [4-8]. Indeed, some species can maintain relatively stable basal energy requirements over broad temperature ranges. For example, in Atlantic halibut (Hippoglossus hippoglossus), SMR was indistinguishable between 10°C and 16°C following 14 weeks acclimation [9]. Thermal acclimation capacity is, therefore, increasingly considered as a key feature determining resilience to climate change [7,10–13], and plastic responses at the individual level can buffer temperature effects potentially allowing time for genetic adaptation to occur at the species level [14-16]. While it is well known that average ocean temperatures are increasing, recent projections also predict more extreme and frequent heat waves [17-19]. It could, therefore, be argued that in a more thermally variable future, the thermal acclimation rate (i.e. the speed by which an organism can compensate for a given change in environmental temperature and attain a new steady state) will become more important. Rapidly acclimating species may, therefore, acquire increasingly greater energetic and competitive advantages as sudden changes in temperature may become more frequent and extreme in the future, but so far the time course and rate of metabolic thermal acclimation in ectothermic animals have been explored little.

Thermal acclimation of metabolism involves changes at multiple organizational levels spanning from biochemical and cellular processes to behavioural responses [5,20-25]. At the whole-animal level not only SMR changes but also there are changes in other metabolic traits affecting the energy budget including the maximum metabolic rate (MMR), aerobic scope (AS, the difference between SMR and MMR) and the metabolic costs of feeding (i.e. the specific dynamic action, SDA). During a transient temperature increase such as during an episodic heat wave, the increase in SMR may lead to a reduction in AS unless the MMR increases by an equivalent or greater amount. The aerobic metabolic reserve available to process beyond basal requirements is then reduced [26-28]. While thermal effects on AS has been proposed to be a key mechanism determining, e.g. fitness and biogeography of fishes with climate change [2,29], little is known about the chronic effects of temperature on AS [6,30]. In Atlantic halibut acclimated to static temperatures between 6°C and 18°C for 14-16 weeks, AS increased continuously with acclimation temperature up to 18°C, which is near lethal temperatures [9]. However, recent work on the tropical barramundi (Lates calcarifer) showed a somewhat different response. While AS in 29°C acclimated barramundi increased continuously with an acute temperature increase up to 38° C, which is $2-3^{\circ}$ from the upper lethal temperature; five weeks acclimation at 38°C resulted in a significantly reduced AS that was not different from 29°C acclimated fish [31]. While these studies suggest that AS can be drastically modulated with thermal acclimation, there appears to be considerable interspecific differences. Moreover, the time course of thermal acclimation of AS remains largely unexplored although this would have important implications for ectotherms in a warmer and more thermally variable future.

The SDA, which are the metabolic processes and subsequent increase in oxygen consumption associated with feeding, must also be accommodated by AS [32,33]. The temperature effects on SDA in fishes are variable, but the postprandial peak metabolic response typically increases with temperature; not only because the SMR may increase but also because the absolute elevation in metabolic rate (i.e. the SDA amplitude) increases [32,34,35]. This presents fish feeding and digesting in a warming environment with a metabolic challenge as digestion may become metabolically constrained, and there may also be increasingly less aerobic capacity available for other fitness-related processes such as locomotion during digestion. Thermal acclimation of the SDA response along with changes in AS could, therefore, have ecologically important implications, but currently information is scant about the thermal acclimation dynamics of SDA and how this relates to changes in AS.

This study explored the temporal aspects of thermal acclimation of SMR, MMR and AS; as well as various components of the SDA in shorthorn sculpin (*Myoxocephalus scorpius*) following a temperature increase from 10°C to 16°C. This species inhabits coastal Atlantic waters throughout much of the Northern Hemisphere and is a typical ambush predatory species that remains motionless on the bottom substratum for extended periods of time [36]. We hypothesized that the thermal acclimation dynamics in this sculpin species would have considerable ecological

relevance as it is probably exposed to significant variations in temperature in its littoral habitat. Furthermore, we predicted that the postprandial metabolism would constitute a large proportion of the available AS given its inactive lifestyle [37], and documented ability to consume large prey items [38,39]. In addition, given the dearth of information on thermal acclimation rate in ectothermic animals, a mathematical model was developed based on field temperature data and our empirical study to evaluate the energetic costs of differing acclimation rates under current and a simulated future ocean temperature scenario.

2. Material and methods

(a) Animals

Shorthorn sculpin (*M. scorpius*) between 80 g and 330 g (mean: 147.2 \pm 9.2 g) were caught by a local fisherman in traps in the vicinity of the Gullmar fjord on the Swedish west coast. Fish were transported to the Department of Biological and Environmental Sciences at the University of Gothenburg, Sweden, and held in approximately 1 m³ tanks supplied with aerated seawater (10°C) from a recirculating system. The animals were fed ad libitum at least once a week with Alaskan pollock (*Theragra chalcogramma*) and occasionally with northern shrimps (*Pandalus borealis*).

(b) Experimental protocols

Fish were anaesthetized in seawater containing MS222 (100 mg l^{-1}) and implanted with a Passive Integrated Transponder tag in the abdominal cavity and marked subcutaneously with Visible Implant Elastomere (Northwest Marine Technology, Washington, DC, USA) for easy visual discrimination. Fish were divided into three experimental groups and left for approximately eight weeks to acclimate to the holding conditions at 10° C.

- (1) In the first group, the dynamics of the temperature acclimation response of SMR and MMR was determined after increasing the temperature to 16°C. This scenario was considered ecologically relevant as these temperatures and magnitude of temperature change are within the range of thermal conditions observed during summer in the Gullmar fjord (http://www.weather.loven.gu.se/data.shtml) where the fish were caught. Measurements were first taken at 10°C and then after one, four and eight weeks of acclimation to 16°C. During measurements, fish were randomly collected from the holding tank and MMR was measured after 5 min of encouraged exhaustive exercise in a separate tank, a treatment that rendered the fish completely unresponsive to physical stimuli at the end of the protocol. The fish was then quickly placed in the respirometer (see below for details) and the highest recorded value was considered to represent MMR. The SMR was then recorded after at least 24 h in the respirometer as the mean over 2 h when the metabolic rate had declined and plateaued [30].
- (2) In the second group, the temperature acclimation response of the postprandial metabolic response (i.e. SDA) was monitored after transfer to 16°C as outlined above. Measurements were first taken at 10°C and then after one, four and eight weeks of acclimation to 16°C. Fish were randomly collected from the holding tank, placed in a respirometer and allowed a recovery period of 24 h. The SDA response was determined by gavage feeding the fish a standardized ration of cut pieces of fish meat (i.e. *T. chalcogramma*) amounting to 5% of the body mass. The oxygen consumption rate was then measured until the metabolic rate had returned to baseline.

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(3) A third control group was maintained at 10°C for the duration of the experimental period to account for, for example, seasonal variations in metabolic rates.

The two experimental groups (i.e. groups 1 and 2) were kept collectively in one tank. The control group (group 3) was kept in a nearby identical tank connected to the same recirculating water system.

(c) Respirometry

Eight custom-made round PVC respirometers (total volume: 3.1 l), submerged in an outer 240 l tank and supplied with a continuous flow of aerated seawater, was used to measure oxygen consumption rate (MO₂) by intermittent-flow respirometry [30]. Circulation of water through the respirometers was achieved by submersible pumps, one mixing the water inside the respirometer and one controlled by an automatic time relay to close the respirometer for 15-20 min during MO₂ measurements. The duration of the closed state was adjusted to avoid water oxygen tension falling below 65% air saturation. Oxygen levels were measured using Oxi 340i oxygen meters (WTW, Weilheim, Germany) placed in line with the mixing pumps. Analogue signals from the oxygen metres were stored digitally using a PowerLab system connected to a computer running LABCHART v. 7.2.2 software (ADInstruments, Castle Hill, Australia). After each experiment, respirometers were cleaned and rinsed with hot water to remove any microbial growth. Background oxygen consumption was checked regularly in empty respirometers and was always negligible.

(d) Calculations

MO₂ was calculated as

$$\mathrm{MO}_{2} = \frac{\Delta[\mathrm{O}_{2}] \times \alpha_{\mathrm{O}_{2}} \times \upsilon}{m \times t}$$

where $\Delta[O_2]$ is the difference in oxygen concentration (%) calculated from the slope of the resultant linear regression; α_{O_2} is oxygen content (mg l⁻¹) of water; *v* is volume of the closed respirometer excluding the volume of the fish, *m* is body mass of the fish and *t* is time. MO₂ was adjusted to represent a 100 g fish and normalized to body mass (i.e. mg O₂ kg⁻¹ h⁻¹) according to [40]:

$$MO_{2(100)} = MO_2 \times \left(\frac{m}{100}\right)^{(1-A)}$$

where $MO_{2(100)}$ is oxygen consumption of a 100 g fish and MO_2 the oxygen consumption of a fish with a body mass of *m*, as calculated above. *A* is the mass exponent (0.8). AS was calculated as the difference between MMR and SMR.

Postprandial metabolic rate was analysed as the average of three MO₂ recordings every 10 h. The following components of the SDA response was calculated for individual fish: (i) SDA, the total amount of oxygen consumed, calculated as the integrated area under the curve using a fitted polynomial trend line; (ii) SDA duration, the duration of elevated MO2 until metabolism had returned to within 10% of the initial MO_2 ; (iii) peak postprandial MO₂, the maximal postprandial oxygen consumption rate; (iv) SDA amplitude, the difference between peak postprandial MO2 and SMR; (v) time to peak postprandial MO₂, the time from feeding until peak postprandial MO₂; (vi) SDA coefficient, the total energy used in relation to the energetic content of the meal, this was calculated by dividing the SDA with the energy content of the meal, using an oxycalorific coefficient of $14.06 \text{ kJ g } O_2^{-1}$ [41]; and (vii) the SDA quotient, the ratio between the SDA and the total SMR. In addition, we estimated the remaining AS during peak SDA [42]. This was done by subtracting peak postprandial MO₂ in individual fish



Figure 1. Thermal acclimation dynamics of metabolic rate following transfer from 10°C to 16°C in shorthorn sculpin (*M. scorpius*). (*a*) SMR (filled bars) and MMR (open bars) and (*b*) AS. Measurements were taken on 10°C acclimated animals prior to transfer (10°C, blue bars) and after one, four and eight weeks acclimation at 16°C (red bars). Variables are reported as means \pm s.e.m. (n = 11-12). Q_{10} values for the change in SMR from 10°C are presented for each column. Asterisks denote significant (* $p \leq 0.05$ and ** $p \leq 0.01$) differences from the respective value at 10°C. (Online version in colour.)

from the mean MMR of unfed fish, assuming that MMR does not change with feeding.

(e) Modelling energetic consequences of differing acclimation rates

As sculpins were found to show complete thermal acclimation of SMR over the temperature range examined (figure 1), a mathematical model based on our empirical data was constructed to estimate the accumulated excess energetic cost and duration of the acclimation process. This was done by fitting the SMR data of sculpins acclimating to 16°C for one, four and eight weeks (figure 1) to an exponential decay equation:

$f(t) = a(1-b)^t,$

where f(t) represents the temperature-induced change in SMR, a is the estimated increase in SMR at time 0, b the exponential decay coefficient (percentage per day) and t is time. From this model, we calculated: (i) acclimation time as the time in days taken to re-establish SMR within 10% of SMR at 10°C, and (ii) accumulated energetic cost during acclimation (in kilojoules) by integrating the area under the curve of the decay function. Data were normalized to a 100 g fish with oxygen consumption converted to energy using an oxycalorific coefficient of 14.06 kJ g O_2^{-1} [41]. We then modelled how a doubled acclimation rate would affect these variables by doubling the decay coefficient in the model (i.e. $b \times 2$). While the temperature increase of 6°C used in the empirical part of this study corresponds well with current high summer temperatures and thermal fluctuations (see Experimental protocols section), we also evaluated the implications of a hypothetical future ocean **Table 1.** Temperature effects on postprandial metabolism in shorthorn sculpin (*M. scorpius*). (Values are means \pm s.e.m. (n = 3-7). Asterisks denote statistically significant difference from 10°C. For further details see Material and methods section.)

	10°C	16°C		
		1 week	4 weeks	8 weeks
SDA (g $O_2 \text{ kg}^{-1}$)	2.1 ± 0.4	2.0 <u>+</u> 0.4	2.7 <u>+</u> 0.6	1.4 ± 0.2
SDA duration (h)	67 <u>+</u> 3	57 <u>+</u> 5*	66 <u>+</u> 3	67 <u>+</u> 3
peak postprandial MO_2 (mg O_2 kg ⁻¹ h ⁻¹)	86.0 <u>+</u> 8.2	131.6 ± 8.6*	120.9 <u>+</u> 17.4*	86.1 <u>+</u> 7.2
SDA amplitude (mg $0_2 \text{ kg}^{-1} \text{ h}^{-1}$)	48.3 <u>+</u> 7.1	59.7 <u>+</u> 7.1*	69.5 <u>+</u> 14.5*	44.4 <u>+</u> 5.6
time to peak postprandial MO_2 (h)	21.4 ± 14.4	18.3 <u>+</u> 3.1	20.0 ± 2.2	23.3 <u>+</u> 3.3
SDA coefficient (%)	9.0 <u>+</u> 1.6	8.8 <u>+</u> 1.9	11.8 <u>+</u> 2.7	6.4 <u>+</u> 1.0
SDA quotient (%)	41.7 <u>+</u> 5.5	27.9 <u>+</u> 4.8	41.4 <u>+</u> 5.5	34.0 <u>+</u> 7.0

scenario with greater fluctuations in temperature extremes (e.g. as caused by a more pronounced episodic heat wave, e.g. [19]). This was performed by estimating the energetic consequences of a 50% greater temperature increase (i.e. $\Delta 9^{\circ}$ C), using a Q_{10} of 2.7 for SMR as obtained one week after transfer from 10°C to 16°C (figure 1).

(f) Statistical analysis

Changes in metabolic rates were statistically analysed using oneway ANOVA or one-way rm-ANOVA followed by Dunnet's post hoc tests to identify time points that were significantly different from the initial value at 10°C. Unpaired student's *t*-tests were used to compare specific time points between experimental groups were such comparisons were considered relevant. Assumptions of normality and homogeneous variance were validated using the Kolmogorov–Smirnov test and the Bartlett's test, respectively. All values are reported as means \pm s.e.m. and $p \leq 0.05$ was considered statistically significant.

3. Results

(a) Thermal acclimation effects on standard metabolic

rate, maximum metabolic rate and aerobic scope

Fish typically remained completely motionless in the respirometers, and so the recorded baseline values closely approximate SMR for this species. SMR in 10°C acclimated fish was $39.2 \pm 3.8 \text{ mg} \text{ O}_2 \text{ kg}^{-1} \text{ h}^{-1}$. One week after the temperature was increased to 16°C, SMR increased to $71.2 \pm 6.8 \text{ mg} \text{ O}_2 \text{ kg}^{-1} \text{ h}^{-1}$, reflecting a Q_{10} of 2.7 (figure 1a). Over the next eight weeks, SMR gradually decreased such that after eight weeks SMR was 40.2 ± 8.7 mg $O_2\,kg^{-1}\,h^{-1}\!,$ which was not significantly different from SMR at 10° C and reflecting a Q_{10} of 1.0. The initial MMR at 10° C was 142.3 ± 9.2 mg O₂ kg⁻¹ h⁻¹. MMR was not significantly different from the value at 10°C after one and four weeks at 16°C, but after eight weeks, it was significantly reduced to $114.9 \pm 6.8 \text{ mg O}_2 \text{ kg}^{-1} \text{ h}^{-1}$ (figure 1*a*). The combined effects on SMR and MMR resulted in a significantly reduced AS from $103.3 \pm 7.3 \text{ mg} \text{ O}_2 \text{ kg}^{-1} \text{ h}^{-1}$ at 10°C to $56.4 \pm 6.4 \text{ mg} \text{ O}_2 \text{ kg}^{-1} \text{ h}^{-1}$ after one week at 16°C (figure 1b), which is only 55% of the value at 10°C. Over the eightweek acclimation to 16°C, AS partly recovered along with the reduction in SMR and increased to 74.7 ± 5.9 mg $O_2 \text{ kg}^{-1} \text{ h}^{-1}$. However, AS was still significantly lower than

the value at 10° C. There were no significant changes in metabolic rates in the control group kept at 10° C (figure 1; electronic supplementary material).

(b) Thermal acclimation of specific dynamic action

At 10°C, feeding resulted in a gradual increase in oxygen consumption, reaching a postprandial peak of $86.0 \pm 8.2 \text{ mg}$ $O_2 \text{ kg}^{-1} \text{ h}^{-1}$ with an amplitude of $48.3 \pm 7.1 \text{ mg} O_2 \text{ kg}^{-1} \text{ h}^{-1}$. The total SDA amounted to $2.1 \pm 0.4 \text{ g} O_2 \text{ kg}^{-1}$ with a duration of $67 \pm 3 \text{ h}$ (table 1). At the peak SDA response, metabolic rate was still significantly lower than MMR in unfed fish at 10°C (figure 2*a*), resulting in an estimated AS during peak SDA of $55.2 \pm 7.0 \text{ mg} O_2 \text{ kg}^{-1} \text{ h}^{-1}$ (figure 3).

One week after the increase to 16°C, the peak postprandial MO₂ reached a significantly higher value of $131.6 \pm$ 8.6 mg $O_2 kg^{-1} h^{-1}$, whereas the SDA duration was reduced to 57 \pm 5 h. The higher SDA peak was mainly caused by the higher SMR because the SDA and the SDA amplitude were not different from 10°C (table 1). In fact, the peak SDA response was now equivalent to MMR in unfed fish (figure 2b), and the estimated AS during peak SDA was close to zero (figure 3). After four weeks acclimation to 16°C, the peak postprandial MO₂ response was still significantly elevated compared with the response at 10°C, mainly because the SDA amplitude was significantly increased (table 1). Although the SDA duration and the SDA were unchanged relative to fish at 10°C, there was no difference between MMR in unfed fish and the peak SDA (figure 2c and table 1), which meant that the estimated AS during digestion remained markedly depressed (figure 3). However, after eight weeks at 16°C, the metabolic response to feeding was more similar to that in 10° C acclimated fish (figure 2*d*). The SDA amplitude was no longer significantly elevated and the peak postprandial response was not different from the response at 10°C. The estimated AS during peak SDA was, therefore, restored to 55% of the 10°C value $(30.5 \pm 7.2 \text{ mg O}_2 \text{ kg}^{-1} \text{ h}^{-1}$, figure 3). The SDA coefficient and quotient did not change with thermal acclimation.

(c) Model of metabolic acclimation rates in future oceans

When fitting SMR data from this study to the thermal acclimation model, a decay coefficient of 0.0125 (i.e. 12.5%



Figure 2. Thermal acclimation dynamics of the postprandial metabolic response (i.e. the SDA response) following transfer from 10°C to 16°C in shorthorn sculpin (*M. scorpius*). Fish were fed with a ration of fish meat corresponding to 5% of the total body mass at vertical arrow. Measurements were taken on 10°C acclimated animals prior to thermal transfer (*a*, blue symbols) and at one, four and eight weeks of acclimation at 16°C (*b*-*d*, red symbols). Variables are reported as means \pm s.e.m. (*n* = 3-7). The shaded area represents SMR and MMR in unfed fish (*n* = 12, see figure 1). Asterisks denote significant (**p* ≤ 0.05 and ***p* ≤ 0.01) changes from 10°C. Daggers denote significant (*p* ≤ 0.05) difference between the peak postprandial metabolic response and MMR. (Online version in colour.)



Figure 3. Thermal acclimation dynamics of the estimated AS during peak SDA following transfer from 10°C to 16°C in shorthorn sculpin (*M. scorpius*). Variables are reported as means \pm s.e.m., (n = 3-7). Asterisks denote significant ($p \le 0.05$) differences from 10°C. For details see Material and methods section. (Online version in colour.)

decrease day⁻¹) was obtained (figure 4*a*). This acclimation rate resulted in an accumulated excess energetic cost of 20 KJ when acclimating to the 6°C increase (figure 4*a*,*b*); while the energetic cost was more than halved (9 KJ) with a simulated doubled acclimation rate (25% decrease day⁻¹; figure 4*c*,*d*). Similarly, the acclimation times were 21 and 10 days, respectively (figure 4*a*,*c*).

A future ocean temperature scenario with a 50% greater temperature increase (9°C) resulted in increased energetic costs during acclimation. With the normal acclimation rate, the excess energetic cost during acclimation increased to 33 KJ, while the cost was 15 KJ with the doubled acclimation rate. Furthermore, when comparing metabolic costs of the future temperature scenario with current thermal conditions, the excess energetic cost was 13 KJ greater with the normal acclimation rate, whereas the difference was only 6 KJ with the doubled acclimation rate (figure 4*b*,*d*). Similarly, the future temperature scenario extended the acclimation time by 4 days with the normal acclimation rate, but only by 2 days with the doubled acclimation rate (figure 4*a*,*c*). The overall differences between the two temperature scenarios ($\Delta 6^{\circ}$ C versus $\Delta 9^{\circ}$ C), therefore, decreased markedly with the faster acclimation rate.

4. Discussion

(a) Thermal metabolic compensation in unfed fish

This study revealed that the shorthorn sculpin (*M. scorpius*) has the capacity to fully compensate for the acute thermal effects on SMR following a temperature increase from 10°C to 16°C. After eight weeks acclimation to 16°C, SMR was not statistically different from the value at 10°C, and the Q_{10} value was 1.0, indicating complete thermal compensation of SMR (i.e. type 2 compensation; [8]). This finding is qualitatively consistent with a recent long-term study on Atlantic halibut (*H. hippoglossus*) [9]. This also gives some hope that macrophysiological predictions on dramatic reductions in fish body size resulting from thermal effects on metabolism with current warming trends may not be as severe because such predictions typically employ SMR Q_{10} values above 2 [43].

The MMR was initially unchanged after transfer to 16° C, and then declined slightly. The greater reduction in SMR, therefore, resulted in considerable thermal compensation of AS. Given that maintenance of a high AS is generally



Figure 4. Effects of different acclimation rates in future ocean temperature scenarios. SMR (*a*,*c*) and the accumulated excess energetic cost during acclimation (*b*,*d*) following temperature increase by 6°C (current, red) or 9°C (future, black). Fish with an acclimation rate of 12.5% decrease day⁻¹ in (*a*,*b*), and fish with a doubled acclimation rate of 25% decrease day⁻¹ in (*c*,*d*). Hatched bars in (*a*) are empirical SMR data (figure 1) used to fit the exponential decay function. Arrows mark the time to 90% thermal compensation of SMR. Δ_{cost} in (*b*,*d*) represents the difference in the accumulated excess energetic cost between temperature scenarios. (Online version in colour.)

considered adaptive [29,44], it is perhaps immediately less obvious why MMR was not increased or at least maintained with thermal acclimation. This suggests that there are either costs associated with maintaining a high MMR, or perhaps the reduction in MMR is simply an inevitable consequence of the reduced SMR. Nonetheless, the small decrease in MMR with warm acclimation is partly consistent with a recent study on the tropical barramundi where both MMR and AS were markedly reduced with thermal acclimation following an initial increase with acute temperature increase [31]. However, the reduced AS observed in this previous study differs from the present study where AS was partly restored during acclimation. In the study on Atlantic halibut, AS increased continuously with increasing acclimation temperature, but the acute thermal effects on AS were not examined [9]. Unless these sculpins are able to migrate into cooler (e.g. deeper) waters, the reduction in AS initially observed here at the higher temperature would probably have negative fitness consequences in nature; e.g. by constraining feed processing and impair predator avoidance by reducing the scope for activity [26,44]. Even so, it appears increasingly clear that the chronic thermal effects on AS may be quite variable and differ considerably between species and possibly with different heating regimes. Compensatory thermal acclimation responses on AS should therefore be better integrated when forecasting the effects of climate change on ectothermic animals [6,7,30]. Yet, detailed knowledge about the ecological ramifications of AS in wild animals remains limited [30].

The SMR for *M. scorpius* reported here at 10°C and after eight weeks at 16° C (approx. 39 mg O_2 kg⁻¹ h⁻¹) are lower than a previous study on the same species from the North sea acclimated to 5–6°C and measured at 7°C (approx. 48 mg $O_2 kg^{-1} h^{-1}$) [45], but slightly higher than conspecifics acclimated and measured at 15°C (approx. 31 mg O_2 kg⁻¹ h⁻¹) [39]. Even so, when revisiting a recent study on the same species from Greenland waters that were field-acclimated to approximately 9°C [36], SMR reported here is nearly half of the values in the Greenland study ($80.4 \pm 8.9 \text{ mg O}_2 \text{ kg}^{-1} \text{ h}^{-1}$). This is interesting because these sculpins were examined by the same researchers using a nearly identical experimental set-up. While this may suggest that the sculpins from the Arctic population had not yet fully acclimated to the unusually warm Arctic summer conditions in 2009 [36], it seems equally possible that the Greenland population had an elevated metabolic thermal sensitivity resulting from, for example, genetic adaptation and/or non-genetic (e.g. epigenetic) effects imposed by the frigid Arctic conditions. This finding is consistent with a recent meta-analysis of fish species across latitudes showing that populations from high latitudes have relatively higher SMR and aerobic capacities [46].

(b) Thermal acclimation of the postprandial metabolic response

The peak SDA response typically increases while the SDA duration decreases with temperature in ectotherms [32,34,47]. Accordingly, after one week at 16°C a significantly

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higher peak postprandial metabolism was observed in shorthorn sculpin. This increase was entirely caused by the increase in SMR because the SDA and the SDA amplitude were unchanged, although the SDA duration decreased (table 1). As the estimated AS during peak SDA was initially close to zero at 16°C, it seems likely that digestion was metabolically constrained at this point. In fact, after four weeks at 16°C, when SMR had been partly thermally compensated, feeding resulted in greater SDA amplitude; but the estimated postprandial AS was still close to zero, suggesting that SDA was still metabolically constrained. The initially reduced ability to digest a large prey item following an acute temperature increase would of course be physiologically and energetically challenging for these sculpins in nature, not least because the elevated SMR requires an increased energy intake to maintain energy balance. Furthermore, peak postprandial metabolism typically increases with meal size [32], and so digestion of even larger meals would probably be increasingly constrained by the reduced AS with warming. The estimated reduction of postprandial AS also indicate that the capacity for other metabolically demanding activities such as predatory avoidance and inter- and intra-specific competition will be severely compromised during digestion at the higher temperature.

Nonetheless, following long-term acclimation to 16°C these sculpins clearly had the capacity to thermally compensate their SMR such that AS was partly restored and the SDA was no longer metabolically constrained. Even so, a potential caveat in these calculations is that feeding may have additive metabolic effects in some fish species resulting in an elevated MMR postprandial [34,47-49], although this is not the case in all species [50,51]. It is, therefore, possible that we may have underestimated postprandial AS, although it remains uncertain how temperature acclimation affected this. A recent study showed that MMR was higher postprandial in three cyprinids acclimated to 15°C, but this additive effect partly disappeared after acclimation to 25°C where only one species showed a clear additive effect [34]. This further highlights the complex relationship between temperature and feeding energetics in wild animals.

(c) Implications of thermal acclimation rate in future oceans

Although comparative data are scarce, it appears that the thermal compensation of SMR in shorthorn sculpin in this study is a relatively slow process. For example, in rainbow trout (*Oncorhynchus mykiss*) taken acutely from 8°C to 18°C, SMR only showed partial thermal compensation, but a new steady state was established within approximately 3 days [52]. In the Antarctic bald notothen (*Pagothenia borchgrevinki*), there was no statistically significant difference in SMR

between animals at -1° C and 4° C after 9 days acclimation to the higher temperature [53].

While the capacity for thermal acclimation is generally regarded as a key determinant of resilience to climate change [12,14], this study emphasizes that the acclimation rate should also be incorporated into future models. Our study suggests that a slow acclimation rate may be increasingly energetically disadvantageous in a future with more pronounced episodic heat waves [19]. This is because the accumulated excess energetic cost during thermal acclimation is larger for a slowly acclimating species, and this difference becomes more pronounced with greater thermal variability leading to greater energy expenditure. In addition, the increase in acclimation time is less pronounced in rapidly acclimating species. This does not only affect the direct energetic costs, but also other fitness-related traits such as AS and SDA that are inherently dependent on the thermal resetting of SMR. For simplicity, we assumed complete thermal compensation in both thermal scenarios tested in the model, as the main purpose was to explore the effects of differences in the rate of acclimation. However, in reality, it is possible that species may be unable to fully compensate if thermal variability increases and this would off course exacerbate the metabolic consequences. While the ecological implications of these differences in acclimation rate are largely unknown, they can potentially lead to competitive imbalances between today coexisting species that have similar acclimation capacity but different acclimation rates. Even so, individuals with relatively faster acclimation rates will probably pay a smaller energetic prize and experience competitive advantages in future oceans with more variable and extreme thermal conditions. It could then be speculated that if global change progresses, a likely evolutionary outcome would be that natural selection will favour phenotypes with faster thermal acclimation rates.

Given the dearth of information on the dynamics and capacity of metabolic thermal acclimation in ectothermic animals, future studies should more thoroughly explore the time course of thermal acclimation and the underlying changes of critical physiological and biochemical processes. Particularly, in temperate species that may have evolved to compensate physiologically for large but relatively slow seasonal temperature changes, future 'winners' may turn out to not only be those that *can* acclimate, but also perhaps more importantly, those who can acclimate fast.

Ethics statement. All experiments were covered by ethical permit 143-2010 from the regional animal ethics committee.

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